Invertebrates of temporary waters in gnammas on granite outcrops in Western Australia

I A E Bayly
Department of Biological Sciences,
Monash University, Clayton VIC 3168

Abstract

Thirty-six flooded gnammas (rock pools), distributed between 17 different granite outcrops, were each sampled once only during the winter of 1990. The water of most pools was acidic (pH < 7.0) and of low conductivity (K < 200 µS cm⁻¹). A total of 88 invertebrate taxa were found with Boeckella opaqua (Copepoda; 19 occurrences), Cypretta baylyi (Ostracoda; 17) and Neothrix armata (Cladocera; 14) most common. A high proportion of the Crustacea consisted of species endemic to Western Australia. Six new species of Cypricercus (Ostracoda) were discovered. The mean number of taxa was 8.2 per pool. There was a highly significant positive correlation between species richness and the logarithm of both the pool volume and pool area. The nature of the fauna is reviewed within the framework of the type of adaptations employed by the animals for tolerating or avoiding the dry phase.

Introduction

The word “gnamma” is of Aboriginal origin and refers to a depression that has been weathered out of the surface of an outcrop of bare rock; some overseas workers (e.g. Smith 1941) have referred to these depressions as “weather pits”. Gnammas, or weather pits, are commonly found on granite outcrops and especially on the top of domed inselbergs. Despite the widespread occurrence of gnammas in Australia, the aquatic biota of rain-filled gnammas has received remarkably little attention. This neglect might be partly explicable on the basis that, because gnammas are decidedly at the small end of the size spectrum of lentic waters (pool-pond-lake), they have been regarded as being of little interest or consequence to aquatic ecologists. In fact, small size confers on gnammas a number of methodological advantages, such as ease of sampling and experimental manipulation, as subjects for an ecologist. Sheldon (1984), for example, draws attention to the good potential of rock pools for comparative and experimental studies of the aerial colonization dynamics of adult aquatic insects. This claim for gnammas, along perhaps with phytotelma (pockets of water held in plants such as the Albany Pitcher Plant; Bayly 1984), as being quintessential aquatic microcosms, is appealing. It may be claimed with some confidence, therefore, that the potential of studies of gnammas for illustrating ecological principles is just as great, if not greater, than that of large lakes.

Materials and Methods

This account is based on collections made from 36 gnammas, distributed between 17 different granite outcrops, during the winter of 1990. Thirteen of the 17 outcrops were in the form of domical inselbergs while the remainder were small flat surfaces. With one exception, the gnammas were shallow, flat-bottomed pan-gnammas sensu Twidale & Corbin (1963). The exception was a deep pit-gnamma located on War Rock.
aperture was 150 µm, and preserved in either 10% formalin or 90% ethanol.

The product of the maximum length and maximum width of a pool was used to approximate (consistently overestimate) the true areas of the pools. Likewise, the product of the maximum length, maximum width and maximum depth was used to approximate (consistently overestimate) the true volumes of the pools. The approximated areas and volumes of the pools were log-transformed for correlation calculations.

Results

Data on the location and physical dimensions of the 36 pools as well as their water chemistry are presented in Table 1. With one exception, pH was within the range 4.6–7.9, but for 31 of the pools it was less than 7.0. With three exceptions, the conductivity (K₂₅) was less than 1000 µS cm⁻¹, and for the 33 pools whose conductivity was less than this value the mean was 147 µS cm⁻¹.

The results of taxonomic studies are summarized in Table 2. A total of 88 taxa were recorded and the mean number was 8.2 per gnamma. Branchiopod crustaceans were found in 18 pools, with *Limnadia* the most frequently occurring taxon. Cladocera occurred in 26 pools, with *Neothrix armata* the commonest species. Ostracods occurred in all but one gnamma and were the only animals collected from four gnammas. The commonest ostracod was *Cypretta baylyi*. At least six new species of *Cypricercus* were recorded. Insects were found in 19 pools, with *Anisops thienemanni* the commonest species. Pools 16 and 21 yielded a disproportionately high number of insect species. Oribatid mites were restricted to two small coastal pools.

There is a highly significant, positive correlation between species richness and the logarithm of approximated pool volume (Fig 1; Pearson r = 0.595, n = 36, P<0.001). If one outlier (two taxa from pool 19 with log volume 4.78) is removed from the data set, then the

### Table 1

Table 1: Physico-chemical features of Western Australian granite rock-pools sampled June-August 1990.

<table>
<thead>
<tr>
<th>Locality number and name</th>
<th>Location (lat S, long E)</th>
<th>Sampling date</th>
<th>Maximum depth (cm)</th>
<th>Max. length x max. width (m²)</th>
<th>pH</th>
<th>Conductivity (K₂₅; µS cm⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Coragina Rock (a)</td>
<td>32°55', 123°30'</td>
<td>24.vi.1990</td>
<td>25</td>
<td>16 x 7</td>
<td>8.7</td>
<td>91</td>
</tr>
<tr>
<td>2. Coragina Rock (b)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>6</td>
<td>3 x 2</td>
<td>6.4</td>
<td>59</td>
</tr>
<tr>
<td>3. Mt Madden (a)</td>
<td>33°14', 119°50'</td>
<td>26.vi.1990</td>
<td>30</td>
<td>30 x 10</td>
<td>6.3</td>
<td>74</td>
</tr>
<tr>
<td>4. Mt Madden (b)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>10</td>
<td>12 x 2.5</td>
<td>6.0</td>
<td>96</td>
</tr>
<tr>
<td>5. Mt Madden (c)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>15</td>
<td>5 x 2.5</td>
<td>6.1</td>
<td>38</td>
</tr>
<tr>
<td>6. Mt Madden (d)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>20</td>
<td>7 x 5</td>
<td>6.0</td>
<td>61</td>
</tr>
<tr>
<td>7. Cable Beach (a)</td>
<td>35°07', 117°54'</td>
<td>27.vi.1990</td>
<td>4</td>
<td>1 x 0.3</td>
<td>7.1</td>
<td>1480</td>
</tr>
<tr>
<td>8. Cable Beach (b)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>2</td>
<td>1 x 0.3</td>
<td>7.0</td>
<td>1490</td>
</tr>
<tr>
<td>9. Frenchman Bay Rd (a)</td>
<td>35°06', 117°57'</td>
<td>27.vi.1990</td>
<td>2</td>
<td>3 x 2</td>
<td>7.3</td>
<td>380</td>
</tr>
<tr>
<td>10. Frenchman Bay Rd (b)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>5</td>
<td>0.8 x 0.2</td>
<td>6.4</td>
<td>457</td>
</tr>
<tr>
<td>11. Muirilup Rock (a)</td>
<td>34°39', 116°15'</td>
<td>2.vii.1990</td>
<td>12</td>
<td>3 x 1.5</td>
<td>4.6</td>
<td>93</td>
</tr>
<tr>
<td>12. Muirilup Rock (b)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>6</td>
<td>2 x 1.5</td>
<td>5.5</td>
<td>101</td>
</tr>
<tr>
<td>13. Cape Leeuwin (a)</td>
<td>34°22', 115°08'</td>
<td>6.vii.1990</td>
<td>2</td>
<td>2.5 x 1.2</td>
<td>6.7</td>
<td>450</td>
</tr>
<tr>
<td>14. Cape Leeuwin (b)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>4</td>
<td>5 x 1.5</td>
<td>6.5</td>
<td>8040</td>
</tr>
<tr>
<td>15. Bilya Rock</td>
<td>29°00', 115°51'</td>
<td>12.vii.1990</td>
<td>11</td>
<td>3 x 2.5</td>
<td>6.1</td>
<td>90</td>
</tr>
<tr>
<td>16. War Rock (a)</td>
<td>29°05', 116°00'</td>
<td>13.vii.1990</td>
<td>22</td>
<td>25 x 9</td>
<td>5.2</td>
<td>55</td>
</tr>
<tr>
<td>17. War Rock (b)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>&gt;100</td>
<td>6 x 2</td>
<td>7.9</td>
<td>798</td>
</tr>
<tr>
<td>18. War Rock (c)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>12</td>
<td>4 x 2</td>
<td>6.1</td>
<td>38</td>
</tr>
<tr>
<td>20. Bunijl Rock (b)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>9</td>
<td>8 x 5</td>
<td>6.1</td>
<td>34</td>
</tr>
<tr>
<td>22. Petrudor Rocks (b)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>6</td>
<td>3 x 1.5</td>
<td>6.0</td>
<td>186</td>
</tr>
<tr>
<td>23. Petrudor Rocks (c)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>19</td>
<td>6 x 3</td>
<td>6.2</td>
<td>190</td>
</tr>
<tr>
<td>24. Elachbutting (a)</td>
<td>30°36', 118°37'</td>
<td>15.viii.1990</td>
<td>10</td>
<td>6 x 2</td>
<td>5.1</td>
<td>164</td>
</tr>
<tr>
<td>25. Elachbutting (b)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>19</td>
<td>6 x 5</td>
<td>5.7</td>
<td>162</td>
</tr>
<tr>
<td>26. Sanford Rock (a)</td>
<td>31°14', 118°46'</td>
<td>15.viii.1990</td>
<td>26</td>
<td>8 x 3</td>
<td>6.3</td>
<td>102</td>
</tr>
<tr>
<td>27. Sanford Rock (b)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>12</td>
<td>2 x 2</td>
<td>6.7</td>
<td>88</td>
</tr>
<tr>
<td>29. Jillbadgie Rocks (b)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>20</td>
<td>13 x 6</td>
<td>6.3</td>
<td>42</td>
</tr>
<tr>
<td>30. Mt Hampton (a)</td>
<td>31°45', 119°04'</td>
<td>16.viii.1990</td>
<td>33</td>
<td>10 x 9</td>
<td>6.3</td>
<td>144</td>
</tr>
<tr>
<td>31. Mt Hampton (b)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>13</td>
<td>4 x 3</td>
<td>6.4</td>
<td>64</td>
</tr>
<tr>
<td>32. Yorkzaine Rock (a)</td>
<td>31°26', 117°31'</td>
<td>26.viii.1990</td>
<td>9</td>
<td>7 x 5</td>
<td>6.4</td>
<td>113</td>
</tr>
<tr>
<td>33. Yorkzaine Rock (b)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>12</td>
<td>4 x 3</td>
<td>6.2</td>
<td>146</td>
</tr>
<tr>
<td>34. King Rocks (a)</td>
<td>32°19', 119°09'</td>
<td>28.viii.1990</td>
<td>20</td>
<td>14 x 10</td>
<td>6.6</td>
<td>105</td>
</tr>
<tr>
<td>35. King Rocks (b)</td>
<td>&quot;</td>
<td>&quot;</td>
<td>30</td>
<td>16 x 8</td>
<td>6.4</td>
<td>91</td>
</tr>
<tr>
<td>36. Wave Rock</td>
<td>32°27', 118°54'</td>
<td>28.viii.1990</td>
<td>12</td>
<td>9 x 3</td>
<td>6.3</td>
<td>113</td>
</tr>
</tbody>
</table>
Table 2
List of taxa and their occurrences.

<table>
<thead>
<tr>
<th>Taxa</th>
<th>Pool (see Table 1)</th>
<th>Total number of occurrences</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>CRUSTACEA: ANOSTRACA</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Branchinella longirostris</td>
<td>Wolf</td>
<td>25</td>
</tr>
<tr>
<td>B. sp</td>
<td></td>
<td>3</td>
</tr>
<tr>
<td><strong>CRUSTACEA: CONCHOSTRACA</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cypricercus sp or spp</td>
<td></td>
<td>6,20,22,26,27,31</td>
</tr>
<tr>
<td>C. n sp</td>
<td></td>
<td>6</td>
</tr>
<tr>
<td>C. n sp 1</td>
<td></td>
<td>2</td>
</tr>
<tr>
<td>C. n sp 2</td>
<td></td>
<td>3,4,5,7,30</td>
</tr>
<tr>
<td>C. n sp 3</td>
<td></td>
<td>3</td>
</tr>
<tr>
<td>C. n sp 4</td>
<td></td>
<td>6,9,13,14,15,16</td>
</tr>
<tr>
<td>C. n sp 5</td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>C. n sp 6</td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>Heterocypris incongruens</td>
<td>(Rahmdor)</td>
<td>1</td>
</tr>
<tr>
<td><strong>CRUSTACEA: CLADOCERA</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Alona cf setuloides</td>
<td></td>
<td>32</td>
</tr>
<tr>
<td>A. n sp</td>
<td></td>
<td>3</td>
</tr>
<tr>
<td>Alonella – excia group</td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>Biapertura – macroa group</td>
<td></td>
<td></td>
</tr>
<tr>
<td>sp1</td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>sp2</td>
<td></td>
<td>11,35</td>
</tr>
<tr>
<td>sp3</td>
<td></td>
<td>5,12,23,25,26,29,30,34,36</td>
</tr>
<tr>
<td>B. rigidicandus Smirnov</td>
<td></td>
<td>6,27,28</td>
</tr>
<tr>
<td>B. n sp</td>
<td></td>
<td>9,11</td>
</tr>
<tr>
<td>Ephemeropus-barroosi group</td>
<td></td>
<td></td>
</tr>
<tr>
<td>sp1</td>
<td></td>
<td>6,34,35</td>
</tr>
<tr>
<td>sp2</td>
<td></td>
<td>11</td>
</tr>
<tr>
<td>Monospilus dorus Smirnov &amp; Timms</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Plicaricirulus alticarinatus Frey</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Plurispina chaesiodes Frey</td>
<td></td>
<td></td>
</tr>
<tr>
<td>P. multiformica Frey</td>
<td></td>
<td>26,25,33</td>
</tr>
<tr>
<td>Rak stagnensis Frey</td>
<td></td>
<td>1,43,27,29,34</td>
</tr>
<tr>
<td>Leberis aenigmata Smirnov</td>
<td></td>
<td>27,29,31</td>
</tr>
<tr>
<td><strong>Other families</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Daphnia jollyi Petkovski</td>
<td></td>
<td>23,26,29,31</td>
</tr>
<tr>
<td>Neothrix armata Gurney</td>
<td>1,3,6,12,15,18,21,</td>
<td></td>
</tr>
<tr>
<td>Macrotobiota breviseta Smirnov</td>
<td></td>
<td></td>
</tr>
<tr>
<td>M. hardingi Petkovski</td>
<td></td>
<td>24,27</td>
</tr>
<tr>
<td>M. indistincta Smirnov</td>
<td></td>
<td>1,2</td>
</tr>
<tr>
<td>M. longiseta Smirnov</td>
<td></td>
<td>2</td>
</tr>
<tr>
<td>Ceriodaphnia sp</td>
<td></td>
<td>1,2,15,18</td>
</tr>
<tr>
<td>Moina sp</td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>CRUSTACEA: COEPODIDA</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Boeckella opaque Fairbridge</td>
<td></td>
<td></td>
</tr>
<tr>
<td>B. triarticulata (Thomson)</td>
<td></td>
<td>17,19,21,23</td>
</tr>
<tr>
<td>Calommaea ampla (Searle)</td>
<td></td>
<td>21</td>
</tr>
<tr>
<td><strong>Cyclopoidea</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mesocylops australiensis</td>
<td>(Sars)</td>
<td>2</td>
</tr>
<tr>
<td>Metacylops sp</td>
<td></td>
<td>3,4</td>
</tr>
<tr>
<td>Microcylops varius (Sars)</td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>Harpacticoida</td>
<td></td>
<td>1</td>
</tr>
<tr>
<td><strong>CRUSTACEA: OSTRACODA</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Alvaeus n sp</td>
<td></td>
<td>16</td>
</tr>
<tr>
<td>Bemnelonia barangaroo De Decker</td>
<td></td>
<td>1,52,25,29</td>
</tr>
<tr>
<td>B. sp</td>
<td></td>
<td>32</td>
</tr>
<tr>
<td>Candonocypris novozelandiae (Baird)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cygretta bayleyi McKenzie</td>
<td></td>
<td>3,4,6,11,12,16,18,20,22,23,</td>
</tr>
<tr>
<td>C. sp</td>
<td></td>
<td>2,10,15,22,25,28,31,32,33,36</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Taxa</th>
<th>Pool (see Table 1)</th>
<th>Total number of occurrences</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cypricercus sp or spp</td>
<td></td>
<td>6,20,22,26,33,34</td>
</tr>
<tr>
<td>C. n sp 1</td>
<td></td>
<td>2</td>
</tr>
<tr>
<td>C. n sp 2</td>
<td></td>
<td>3,4,5,8,30</td>
</tr>
<tr>
<td>C. n sp 3</td>
<td></td>
<td>3,34</td>
</tr>
<tr>
<td>C. n sp 4</td>
<td></td>
<td>6,9,13,14,15,16</td>
</tr>
<tr>
<td>C. n sp 5</td>
<td></td>
<td>30</td>
</tr>
<tr>
<td>C. n sp 6</td>
<td></td>
<td>36</td>
</tr>
<tr>
<td>Heterocypris incongruens</td>
<td>(Rahmdor)</td>
<td>2</td>
</tr>
<tr>
<td>Philodromus amplificus De Deckker</td>
<td></td>
<td>2,10,18,20,23,29,31,32,33</td>
</tr>
<tr>
<td>L. candonites De Deckker</td>
<td></td>
<td>9,12,16,18</td>
</tr>
<tr>
<td>L. sp</td>
<td></td>
<td>4,15,21,24,27,36</td>
</tr>
<tr>
<td>Limnothoea molnayensis Chapman</td>
<td></td>
<td>2,3,7,34,35,36</td>
</tr>
<tr>
<td>L. sp 1</td>
<td></td>
<td>8</td>
</tr>
<tr>
<td>L. n sp 7</td>
<td></td>
<td>13</td>
</tr>
<tr>
<td>L. n sp</td>
<td></td>
<td>32</td>
</tr>
<tr>
<td>Sarscypridopsis aculaeata (Costa)</td>
<td></td>
<td>17,21,23</td>
</tr>
<tr>
<td><strong>ACARI: ORIBATEI</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Scaphoidea sp 1</td>
<td></td>
<td>78</td>
</tr>
<tr>
<td>Trionalaonothrus sp</td>
<td></td>
<td>78</td>
</tr>
<tr>
<td><strong>INSECTA: COLEOPTERA</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Alloplagus bistrigatus (Clark)</td>
<td></td>
<td>16,21,25</td>
</tr>
<tr>
<td>Erotes australis (Erichson)</td>
<td></td>
<td>2</td>
</tr>
<tr>
<td>Lancetes lancedolatus (Clark)</td>
<td></td>
<td>6</td>
</tr>
<tr>
<td>Megaporus howitti (Clark)</td>
<td></td>
<td>3,16</td>
</tr>
<tr>
<td>Necterosoma darwinii (Babington)</td>
<td></td>
<td>11</td>
</tr>
<tr>
<td>Parasterniger Watts</td>
<td></td>
<td>78</td>
</tr>
<tr>
<td>Spermatoporus multilatulatus (Clark)</td>
<td></td>
<td>16,25</td>
</tr>
<tr>
<td>Hydrophilidae</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bersus approximus Fairmaire</td>
<td></td>
<td>16</td>
</tr>
<tr>
<td>Limnozonus zelandicus (Brown)</td>
<td></td>
<td>6</td>
</tr>
<tr>
<td><strong>INSECTA: DIPTERA</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Dasyhelea sp</td>
<td></td>
<td>22,25</td>
</tr>
<tr>
<td>Chironomidae</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Alabaemisia sp</td>
<td></td>
<td>2</td>
</tr>
<tr>
<td>Allostictioides sp</td>
<td></td>
<td>28,30</td>
</tr>
<tr>
<td>Chironomus tepperi Skuse</td>
<td></td>
<td>223</td>
</tr>
<tr>
<td>Dicrotendipes sp</td>
<td></td>
<td>11</td>
</tr>
<tr>
<td>Parabomilla sp</td>
<td></td>
<td>16</td>
</tr>
<tr>
<td>Paratanetipes sp</td>
<td></td>
<td>33</td>
</tr>
<tr>
<td>Orthocladiinae</td>
<td></td>
<td>25,28,30</td>
</tr>
<tr>
<td><strong>INSECTA: HEMIPTERA</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Corixidae</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Agapicorixia parvicipunctata (Hale)</td>
<td></td>
<td>36,16,21,23</td>
</tr>
<tr>
<td>Diaprepocoris personata Hale</td>
<td></td>
<td>11</td>
</tr>
<tr>
<td>Miconocta annae Kirkaldy</td>
<td></td>
<td>16,21</td>
</tr>
<tr>
<td>M. gracilis Hale</td>
<td></td>
<td>2</td>
</tr>
<tr>
<td>M. robusta Hale</td>
<td></td>
<td>4,25,26,36</td>
</tr>
<tr>
<td>Sigara nullula Lansbury</td>
<td></td>
<td>26</td>
</tr>
<tr>
<td><strong>Notonectidae</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anisops buflis Lansbury</td>
<td></td>
<td>56,16</td>
</tr>
<tr>
<td>A. guttatus Hale</td>
<td></td>
<td>21</td>
</tr>
<tr>
<td>A. insperion Kirkaldy</td>
<td></td>
<td>36,17,21</td>
</tr>
<tr>
<td>A. stali Kirkaldy</td>
<td></td>
<td>17,21</td>
</tr>
<tr>
<td>A. thienemmni Lundblad</td>
<td></td>
<td>56,16,17,21,23,25,30</td>
</tr>
</tbody>
</table>

169
correlation is considerably improved ($r = 0.696$, $n = 35$, $P<0.001$). There is also a highly significant positive correlation between species richness (Pearson $r = 0.577$, $n = 36$, $P<0.001$) and the logarithm of approximated pool area (Pearson $r = 0.641$, $n = 35$, $P<0.001$).

**Discussion**

With seven exceptions, the conductivity ($K_{25}$) of the pools (Table 1) was less than 200 µS cm$^{-1}$ — a finding broadly consistent with that of Bayly (1982) for a series of 19 granite rock pools in Western Australia. Six of the seven exceptions were pools with a maritime location (Cable Beach, Frenchman Bay and Cape Leeuwin) and whose high conductivities were clearly caused by the accession of ions from marine salt spray. The remaining exception was a deep pit-gnamma located on War Rock; this gnamma probably rarely loses accumulated ions by means of overflow.

The pH of almost all of the inland pools is acidic, as expected for a granite substratum. The exceptionally high pH recorded for Coragina Rock (a; Table 1) was discussed by Bayly (1992) who attributed it to the use of cement mortar between the exfoliated slabs of granite used to raise the level of the pool.

Although no quantitative data are available, it is noteworthy that granite rock-pools lack the high turbidity that is so characteristic of pools lying on the regolith, especially in arid regions.

The total number of taxa listed in Table 2 is 88. This, however, is an underestimate of the total number of metazoan invertebrate taxa because black planarians (*Bothromesostoma?*) and nematodes were present in several collections but were not identified and are omitted from Table 2. The total of 88 taxa greatly exceeds the 18 invertebrate taxa listed by Bishop (1974) as occurring in a series of shallow pools located on sandstone at Kanangra Walls in the Blue Mountains, New South Wales. The 88 taxa from all 36 pools and the 17 taxa from the richest pool (number 21) may also be compared with the 121 invertebrate taxa reported by Lake et al. (1989) from a single temporary pond (not located on a granitic substratum) in western Victoria that was repeatedly sampled over 7 months.

The animals listed in Table 2 may be classified into four groups according to their adaptations for tolerating or avoiding the dry phase following the system of Wiggins et al. (1980) as modified by Williams (1985).

- **Group A** (Group 1 of Wiggins et al. 1980) consists of permanent residents of the pools that are capable of only passive dispersal. These organisms are dormant during the dry season and typically avoid desiccation as a resistant stage (shelled egg or embryo). All of the Crustacea listed in Table 2 should be assigned to this group, a salient feature of which is the high proportion of species endemic to Western Australia. Such endemics include *Branchinella longirostris*, *Daphnia jollyi*, *Macrothrix hardingi*, *Boeckella opaqua*, *Ilyodromus amplicolis*, *I. candonites*, *Leberis aenigmatosa*, *Plurispina*...
chauliodus and P. multituberculata. This list does not include the new species of Cypriocercus some of which are likely to prove endemic. A distinctive negative feature of this group is the absence of notostracans. Despite their possession of a suite of adaptations (including resistant eggs) for occupancy of small temporary waters, an unknown factor excluded notostracans from granite rock pools studied here. A possible reason is that the concentration of calcium in waters on granite is so low that it is physiologically impossible for Lepidurus or Triops to fabricate their relatively large carapace. The carapaces of Illyodromus and other ostracod taxa living in granite rock-pools contain practically no calcium; instead their carapaces are strengthened by non-calcareous ridges to compensate for this deficiency P De Deckker (pers comm).

- **Group B** (Group II of Wiggins et al. 1980) comprises animals (typically insects) that are capable of active dispersal but which are typically still present during the dry season in a dormant form (quiescent larvae). Allatrossocladus, Paraforniella and Daphnelea are assigned to this group. The ability of the larvae of these taxa to resist desiccation in gnammas was investigated by Jones (1971, 1975).

- **Group C** (Group IV of Wiggins et al. 1980) consists of animals (typically insects) capable of active dispersal and which have a discontinuous presence in the pools; they avoid the dry period altogether by emigrating to permanent waters before the pools dry up. To this group are assigned all the Dystiscidae, Hydrophilidae, Corixidae and Notonectidae listed in Table 2, some 20 species in all.

- **Group D** [Williams’ (1985) replacement for Group III of Wiggins et al.] comprises animals which lack a resistant stage in their life cycle but possess exceptionally good dispersal ability thus allowing a discontinuous presence in the confines of a temporary water basin. A single species, Chironomus tepperi, may be assigned to this group. The ability of this species to rapidly colonize temporary waters has been demonstrated by Maher & Carpenter (1984).

  The biology of the oribatid mites, Chudalupia meridionalis (see Bayly 1992), Scapherameus sp and Trimalaconothrus sp is insufficiently known to allow their assignment within the Wiggins-Williams system.

  Wiggins et al. (1980) mentioned the habit of burrowing into bottom sediments as a behavioural adaptation for avoiding desiccation in temporary pools. While this pattern of behaviour may provide an escape for some animals in most pools, there is little scope for it in most pan-gnammas because of minimal amounts of sediment and the impossibility of burrowing into granite. Deep pit-gnammas left undisturbed for long periods may accumulate a significant amount of sediment and provide an exception.

  Fryer (1985), on the basis of studies in England, concluded that chydorid cladocerans “show an unambiguous preference for large water bodies”. However, this conclusion is of doubtful validity in the Australian context (Frey 1998). Although sampled once only, some gnammas contained four species of Chydoridae and there is presently no evidence that this number is significantly exceeded in large Australian lakes. Timms (1981), for example, sampled the littoral region of Lake Purrumbete, a large (552 ha) freshwater lake in Western Victoria, monthly for a year, but recorded only six chydorid species.

  The finding of a significant positive correlation between species richness and the volume of the pools is consistent with earlier investigations. Ranta (1982) reported a significant correlation between the number of water beetle species and the logarithm of pool volume in a series of 20 coastal (marine supralittoral) rock pools located in the Baltic region. A similar finding for a greater diversity of taxa was made by March & Bass (1995) for a series of six temporary pools located in Oklahoma.

Acknowledgements: I am very grateful to the following for providing taxonomic identifications; the late Professor D G Frey (Chydoridae), P De Deckker (Ostracoda), I Lansbury (Corixidae and Notonectidae), M C Geddes (Anostraca and Conchostraca), D W Morton (Cyclopoida), M Colloff (Oribatei), D H D Edward (Chironomidae and Corethogonidae), J F Lawrence (Hydropilidae) and C H S Watts (Dytiscidae). Because of the importance of ostracods in gnammas, P De Deckker’s task was a particularly onerous one. I thank the Royal Society of Western Australia for inviting me to contribute to the Symposium and for paying my travelling expenses. I am indebted to L Dembinski for the word processing.

References


Jones R E 1974 The effects of size-selective predation and environmental variation on the distribution and abundance of


Smirnov N N & Bayly I A E 1995 New records and further description of *Macrothrix hardingi* Petkovski (Cladocera) from granite pools in Western Australia. Journal of the Royal Society of Western Australia 78:13–14.


Granite outcrops of Western Australia are weathered landforms that occur throughout the state of Western Australia, composed primarily of the rock type granite. All recognised types of this landform can be observed, commonly as bornhardts, but also as inselbergs, castle koppies and Nubbins. Rising abruptly from the surrounding landscape they create a variety of microhabitats for plants, and provide seasonal resources and refuge for a range of animals. These areas thus have rich biodiversity and many Gnammas on granite outcrops: abundance, origins, maintenance, restoration and biodiversity conservation Steve Hopper, Barbara Cook, Noongar elders Gnammas, especially deeper pit-forms which may be semi-permanent, are vital sources of fresh water in semi-arid southwestern Australia. Caring for gnammas is a practice of vital interest. Gnammas are also well known for having a rich invertebrate fauna, as well as some interesting plant genera specialized for an ephemeral wetland habitat. This project aims to review recorded information on gnammas, develop a better understanding of Noongar oral history about them, and devise restoration processes with Noongar elders that both protect and purify the water supply but also care for gnamma invertebrates and plant life.